Microbial analysis of ovine cheese by next generation sequencing

Vladimir Kmet and Dobroslava Bujnakova

Institute of Animal Physiology, Slovak Academy of Sciences, Soltesovej 4, 040 01 Kosice, Slovakia, Email: kmetv@saske.sk

INTRODUCTION

The most common genera of lactic acid bacteria in ovine milk include *Lactococcus*, *Lactobacillus*, *Streptococcus*, and *Enterococcus* (Chebenova-Turcovska et al., 2011; Pangallo et al., 2014). Strains identified in ovine cheese and bryndza cheese by MALDI-TOF analysis belonged to 10 species of non-enterococcal lactic acid bacteria, i.e. *Lactobacillus casei/Lactobacillus paracasei*, *Lactobacillus plantarum*, Lactobacillus rhamnosus, Lactobacillus helveticus, Lactobacillus *delbrueckii*, *Lactobacillus fermentum*, *Lactobacillus brevis*, *Lactococcus lactis*, *Pediococcus pentosaceus*, and *Pediococcus acidilactici* (Kmet and Drugdova, 2012). Moreover Bunesova et al (2014) isolated *Bifidobacterium crudilactis* and *B. animalis* subsp. *lactis* in ovine cheese.

MATERIAL and METHODS

This study aimed to analyse microbiota of the ovine cheese "Slovak Bryndza" and to investigate the presence of Lactobacillus antibiotic resistance, virulence or probiotic genes by pyrosequencing. The 16S rRNA PCR products from ovine cheese and genomes of lactobacilli (*L. plantarum, L. brevis, L. paracasei*) were sequenced using a Roche 454 GS FLX Titanium sequencer platform.

Fig.1 *Lactobacillus* proteins related to antibiotic resistance

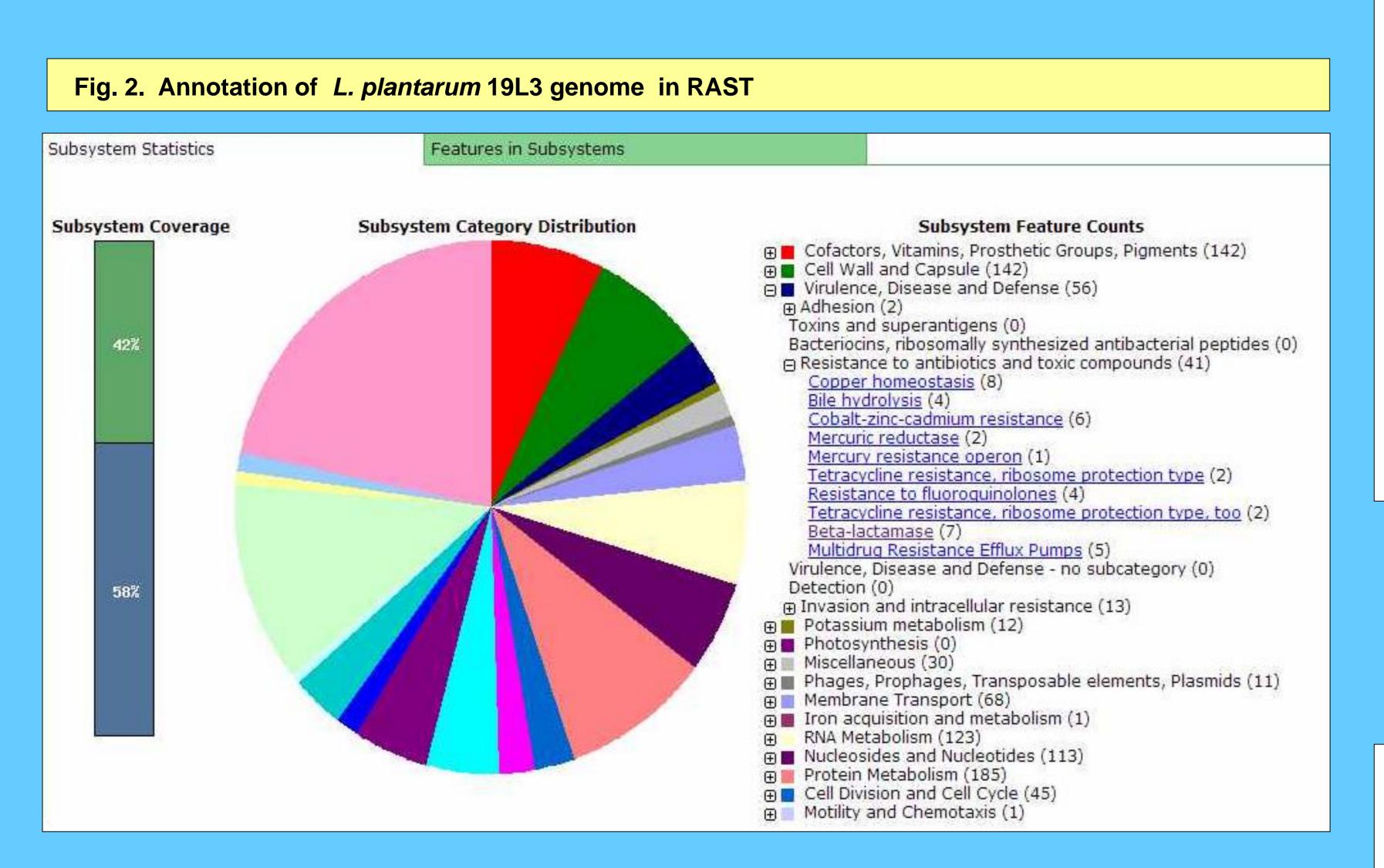
protID	protDesc	pf.rol
PF00905	Penicillin binding protein transpeptidase domain	Serine beta-lactamase-like superfamily
PF03965	Penicillinase repressor	Helix-turn-helix clan
PF00144	Beta-lactamase	Serine beta-lactamase-like superfamily
PF00753	Metallo-beta-lactamase superfamily	NA ?????
PF00204	DNA gyrase B	NA
PF00440	Bacterial regulatory proteins	tetR family
PF01610	Transposase	Ribonuclease H-like superfamily

RESULTS

The V1-V3 regions of the 16S ribosomal DNA were amplified from different ovine cheeses using PCR. In all samples, the microbial populations consisted of *Lactobacillus helveticus*, *Lb. acidophilus*, *Lb. plantarum*, *Lb. rhamnosus*, *Lb. brevis*; *Lactococcus lactis*, *L. raffinolactis*, *L. garviae*; *Enterococcus italicus* and *E. cameliae*; *Streptococcus salivarius*, *St. thermophilus*, *St. caballi* and *St. ferus*.

Laboratory evaluation showed that *L. plantarum* 19L3 was susceptible to antibiotics, according to the ISO 10932 standard

Furthermore, the genomes of selected *Lb. plantarum*, *Lb. brevis*, *Lb. paracasei* were pyrosequenced. The assembly of *L. plantarum* resulted in 203 contigs longer than 1,000 bp (D'Auria *et al.*, 2014).



There were identified probiotic proteins as an alpha amylase (protein ID PF00128), peptidase (PF01433), catalase (PF00199), heat shock protein 33 (PF01430). Nevertheless, there was discrepancy between *Lb. plantarum* ampicillin sensitivity and the presence of serine beta-lactamase like superfamily (Fig. 1). No virulence factors were detected. RAST results (Fig.2) indicated new properties of lactobacilli, which were not occurred by phenotyping testing. EFSA (2012) recommended that probiotic LAB strains should lack acquired antimicrobial resistance and virulence genes detected by PCR or by next generation sequencing.

CONCLUSIONS

Unusual data (antibiotic resistance) from NGS is necessary to verify by PCR. New *Lactobacillus* probiotic proteins were detected.